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Title: Effect of Embedded Polyelectrolyte Chains on Microstructure of Polyacrylamide Hydrogels

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In the present work we have studied the dynamics of chemically cross – linked polyacrylamide (PAAm) hydrogels with and without incorporating polyelectrolyte (pel) chains using dynamic light scattering (DLS) technique. A non – ergodic approach to analyze the intermacromolecular systems suggests stiffening of cross – linked junctions as the concentration of foreign species is increased. Thus the dynamics of parent gels is distinctly different from those in presence of pel chains. To further confirm our observation we have fit the intermediate scattering function to a straight line equation and used it to calculate the diffusion coefficient of the network. It was found that the diffusion coefficient decreased as we increased the added pel chain concentration. The decreased diffusion coefficient indicates an enhanced frozen – in structures in gel matrix.

EFFECT OF EMBEDDED POLYELECTROLYTE CHAINS ON MICROSTRUCTURE OF POLYACRYLAMIDE HYDROGELS

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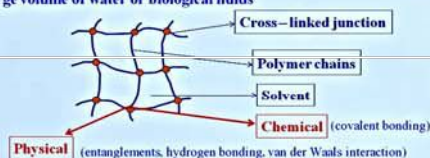
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INTRODUCTION

HYDROGELS:

Three dimensional cross-linked network of polymer chains imbibing large volume of water or biological fluids



- Cross-links – Prevents fluid to flow out
- Trapped fluids – Structural integrity to gel matrix

Structurally flexible, biocompatible, permeable and smart response to external stimuli viz. pH, salt, temperature, electric field

- Generally Gels are known to have a heterogeneous structure and their characteristics is influenced by numerous factors

ELEMENTS INFLUENCING GEL STRUCTURE

- Polymer concentration
 - Cross-linking density
 - Solvent Quality
 - Solution pH
 - Aging of Gels
 - Temperature
- Incorporation of foreign particles – Nano particles, surfactants, Polyelectrolytes

Sodium sulfonated polystyrene (NaPSS) incorporated in Polyacrylamide (PAAM) gel matrix

POLYACRYLAMIDE (PAAM) HYDROGEL:

- Chemically cross-linked gel (monomers are kind of stitched together by cross-linkers which are usually bi- or multi-functional)

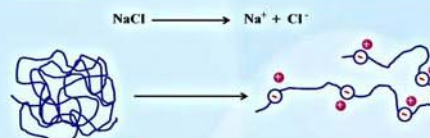
- Monomer: Acrylamide (AAM)
- Cross-linker: N,N' methylene bisacrylamide (BIS)

APPLICATIONS

- Drug treatment
- Ophthalmic operations
- Separation of chemical systems
- Plastic surgery
- Cell culture studies
- Horticulture
- Heavy metal removal
- Sensors
- Water purification
- Biomaterial transport

POLYELECTROLYTES

Ionizable polymer chains - basically dissociates into macroions and counterions

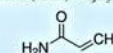


- Polyelectrolytes are of great importance as most of the known biological as well as some of the non-biological polymers are charged.
- Polyelectrolytes are extensively used as transport vehicles for drugs and biomaterials, in water treatment.

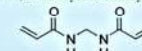
EXPERIMENTAL SECTION

MATERIALS:

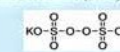
- Monomer - Acrylamide (AAM) - C₃H₅NO



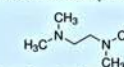
- Cross-linker - N,N' methylene bisacrylamide (BIS) - C₇H₁₀N₂O₂



- Initiator - Potassium Persulfate (KPS) - K₂S₂O₈



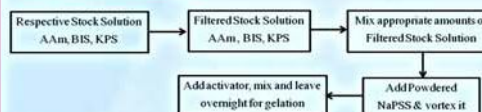
- Activator - N,N,N',N' tetramethylethylenediamine (TEMED)



- Entrapped Polyelectrolyte: Sodium Sulfonated Polystyrene, 1000kDa

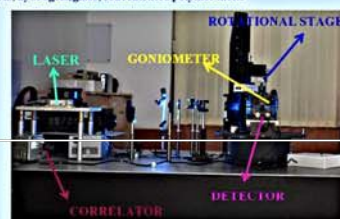


SAMPLE PREPARATION



DYNAMIC LIGHT SCATTERING (DLS)

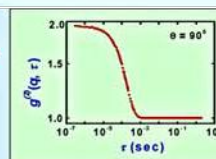
- Used to study dynamics of fluid systems: polymer solutions, colloids, nano particles
- Typically measures time average intensity - intensity correlation function whereas theoretically it is ensemble average
- DLS has recently been used to analyze dynamics in non-ergodic systems viz. hydrogels, glass, concentrated polymer melts



ERGODIC SYSTEM

- All scatterers are free to undergo Brownian excursions
- Therefore in Ergodic systems: $\langle I \rangle_t = \langle I \rangle_\tau$

$$g_2(q, \tau) = 1 + \beta |g_1(q, \tau)|^2$$

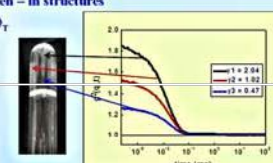


NON-ERGODIC SYSTEM

- All scatterers are not free to move and contribution to scattered electric field: Fluctuating component and Frozen - in structures
- Therefore in Non-Ergodic systems: $\langle I \rangle_t \neq \langle I \rangle_\tau$

$$\sqrt{g^{(2)}(q, \theta)} < 2$$

$$\sqrt{g^{(2)}(q, t)} \text{ varies with position}$$



Fusey & van Megan Method: *Physica A*, 157, 705 (1989)

- Measures $g^{(2)}(q, \tau)$ at one point of the sample
- Measures $\langle I \rangle_t$ at the same point of the sample
- Measure $\langle I \rangle_\tau$ by rotating the sample

By comparing all these values ISF $f(q, t)$ can be calculated

$$g_2^{(2)}(q, t) = 1 + Y^2 [f^2(q, t) - f^2(q, \infty)] + 2Y(1 - Y)[f(q, t) - f(q, \infty)]$$

Intermediate Scattering Function (ISF)

$$f(q, \tau) = \frac{(Y-1)}{Y} + \frac{\sqrt{g_2^{(2)}(q, \tau) - \sigma_i^2}}{Y}$$

$$\text{Long time ISF: } f(q, \infty) = \frac{(Y-1)}{Y} + \frac{\sqrt{1 - \sigma_i^2}}{Y}$$

For diffusive mode: $f(q, t) \propto e^{-Dq^2 t}$

$$f(q, t) = 1 - D_c(q) q^2 t + \dots \quad D_c = Y D_c' \sigma_i^2$$

REFERENCES

1. T. R. Hoare and D. S. Kabane, *Polymer* 49, 1993 - 2007 (2008); 2. A. Tangi, *JPCIS* 4, 951-959 (2014); 3. G. Gerlach and K.F. Arnold, *Hydrogels: sensors and Actuators*, O. Okazaki Springer Series in Chemical Sensors and Biosensors, 2009, Vol. 4, p12 - 4. O. Okazaki and W. Oppermann, *Macromolecules* 40, 3378 - 3387 (2007); 5. C. Ruchon and E. Grisdler, *Macromolecules* 47, 8012 - 8017 (2014); 6. W. Wang and S. A. Saade, *Polymer Journal*, 1 - 9 (2014); 7. P. N. Fusey and W. Van Megan, *Physica A* 157, 705 - 741 (1989); 8. C. Goff and P. G. Righetti, *Electrophoresis* 2, 213 - 219 (1981)

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